

Nutritional characterization of *Detarium microcarpum* Guill. & Perr., seeds used as a soup thickener in Bayelsa State.

***Esther O. Akhigbe, Mbosowo Etukudo and Kelechi J. Ezeoke**

Department of Biology, Faculty of Science, Federal University, Otuoke, Bayelsa State, Nigeria.

ABSTRACT:

Background: This research was carried out to determine the nutritional and phytochemical constituent of *Detarium microcarpum* seeds commonly known as 'ofor' used in Otuoke, Bayelsa State and Environs.

Methods: Fresh fruits of *Detarium microcarpum* were collected from secondary forest in Otuoke and identified. Dry ground seeds were analyzed for mineral elements, proximate composition and phytochemical constituent. Mean and standard error were calculated for the data collected.

Results: Data collected revealed that seeds of *D. microcarpum* are a rich source of essential minerals like magnesium (57.20 mg/100 g) and potassium (46.30 mg/100 g), which are vital for health and functionality of human heart. However, the content of manganese was relatively low (0.16 mg/100 g). Values for proximate content showed high carbohydrate content (63.37%), crude proteins (17.21%) and crude ash (2.81%) which make the seeds a source of balance meal. Furthermore, low water content (8.64%) was observed, which is essential for storage. The results obtained for phytochemical analysis revealed the presence of alkaloids (1.50 mg/ 100 g), tannins (2.97 mg/100 g), flavonoids (1.22 mg/100 g), saponins (1.02 mg/100 g), oxalates (0.53 mg/100 g) and phytates (0.74 mg/100 g).

Conclusion: The result obtained in this study demonstrated that seeds of *Detarium microcarpum* are a good source of vital nutrients and bioactive compounds which can enhance planetary health.

Keywords: *Detarium microcarpum*; nutritional component; phytochemicals; proximate constituent.

1. INTRODUCTION

The world's population is growing fast, and sufficient food is needed to create a balance between economic development and poverty eradication for a healthy and sustainable ecosystem [1]. Plants have been used in the past decades for nutritional and medicinal purposes in both developing and developed countries. Majority of the World's populations depend mainly on plant and their products for their survival [2]. Humans consume food for different purposes, either for therapeutic, nutritional or for pleasure. Certain food eaten by humans might have toxic effect on the body [3]. Insufficient consumption of proteins and micronutrient has been identified as the main cause of infant malnourishment and other diet-related diseases in under developed countries of the world [4,5]. There has been a global increase in food supply from increased agricultural yield in the past few decades [6]. Despite the increase in dietary supply, the nutritional value of food we consume is inadequate. Indigenous trees have been used in sub-sahara Africa for the past decades for food, nutrition and traditional medicinal. They are prospect for income generation if their potentials are maximized [7,8,9]. There has been a dearth of information concerning the nutritional and therapeutic component of many native trees, which has resulted in poor documentation in science and often not acknowledged in

*** Corresponding author: Email: akhigbeeo@fuotuoke.edu.ng; Phone: +234 (0) 805 334 3383**

This is an open-access article distributed under the Creative Commons Attribution License, (<http://creativecommons.org/licenses/by/4.0/>) which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.

Akhigbe et al: Nutritional characterization of *Detarium microcarpum* Guill. & Perr., seeds used as a soup thickener in Bayelsa State.

sustainable development strategies for poverty alleviation [10, 11]. As a result, the nutritional and therapeutic capacity of most traditional trees consumed as food are underestimated and underutilized [12]. It is therefore vital to evaluate the nutritional constituent of the food we consume to estimate sufficient nutrient intake for nourishing individuals at all levels [13]. Adequate information will help facilitate the selection of priority tree food species for domestication programs aimed to foster food security, economic advancement, biodiversity management and conservation. [9]. *Detarium microcarpum* seeds commonly known as ‘ofor’ by the Ibos; ‘ogbogbo’ by the Yorubas and ‘taura’ by the Hausas is a legume native to tropical and Sub tropical Africa regions of the world [14, 15,]. *D. microcarpum* belongs to the family leguminosae and sub-family caesalpiniacea with three main species; *D. microcarpum*; *D. senegalense* and *D. hendelotianum* [16]. Flours from ‘ofor’ seeds have been found to be used in most Eastern regions of Nigeria including Imo, Anambra, Akwa Ibom and Ondo States. They are used as thickeners, emulsifiers and flavouring agents in traditional soups due to their gum content (for eating garri, pounded yam or semo and fufu). These gums are hydrocolloids in nature. [17,18]. flours from the seeds of these legumes have been termed ‘unexploited’. The edible fruits of *D. microcarpum* are a rich source of vitamins C, the root stems, bark are medicinal [19]. Foods, fruits, vegetables are vital immune boosters that help the body fight sicknesses [20]. Phytochemicals have proven to be potent immuno-modulators for therapeutic purposes. The phenolic acids, flavonoids and tannins are the major group of plant secondary metabolites exhibiting immuno-modulating activity and they are frequently found in vegetable and fruits. Phytochemicals contain high antioxidant which have anti-inflammatory properties and thus recommended to mitigate hyper-reacted immune responses which damage tissues through oxidative stress [21]. Immuno-modulators have been utilized for the treatment of various allergic diseases including asthma and allergic rhinitis [22]. Hence, dietary approach is a new model recommended to boost immune and reduce risk of many diseases such as COVID-19. [23]. The consumption of these soup thickeners made it necessary for more research to provide information on their mineral, phytochemical compositions and properties of their constituents so as to determine their actual nutritional benefit, health and other medicinal importance. Also the growing need of the Nigerian Populace calls for immediate analysis on the functionality of locally consume foods so as to increase their utilization. The objective of this study is to determine the mineral nutrients, Proximate and phytochemical composition of *D. microcarpum* seeds in order to maximize and develop strategies for sustainable utilization as a dietary resource. *D. microcarpum* is a shrub with an irregular top with a height of 5-10 m. The tree is easily identified by its leaves which is about 15 cm long, paripinnate or imparipinnate [24,25], with translucent ones. The fruit of *D. microcarpum* is a globular, flattened drupe of 3 to 8 cm in diameter. [26], which is made up of three main parts: the epicarp, mesocarp or pulp, which is intermingled with fibers inserted on the stone (edible pulp) and the stone.

2. MATERIALS AND METHODS

2.1. Materials

The materials used in this experiment were sourced from Biology Laboratory, Federal University Otuoke, Bayelsa State, in alliance with the Laboratories in the environ. Analysis of mineral elements was determined using a drying oven, electric blender, porcelain crucible, a muffle furnace, Hot plate, volumetric Flask Water bath, and atomic absorption spectrophotometer. Proximate analysis required the following materials to determine the nutritional constituent of seeds; a beaker, desiccators, Lenton furnace, cotton wool, extraction flask, porous thimble, soxhlet, Whatmann No. 1 filter paper, Water bath and spatula. Gem filter paper, electronic weighing balance, test tubes, Bunsen burner, burette stand, burette, pipette, funnel, weighing bottle, glaze tile, wire guaze, spectrophotometer, chemical balance were used to determine the phytochemical analysis. Sack bags were used collection of plant samples.

2.1.1. Biological materials

D. microcarpum fruits were collected from the secondary forest in Otuoke, Bayelsa State, Nigeria.

2.1.2. Chemicals and reagents

Chemical reagents in the University Laboratories were used while other were purchased. HNO₃/HCL/H₂O, deionized water were needed for determination of mineral element. Petroleum ether (600cm³) was used for analysis of crude fat, 100 cm³ of 1.25% H₂SO₄, 100 cm³ of 1.25% NaOH, 1:1ethanol and acetone were used for crude fibre content. Conc. H₂SO₄, NAOH/NA₂SO₃, 4% boric acid, indicators, HCL were required for determination of crude proteins. 10% acetic acid, absolute ethanol, diethyl ether, n-butanol, sodium chloride, conc. Ammonium hydroxide, for alkaloid. aqueous



methanol for flavonoids, oxalic acid, potassium permanganate solution, 1.0M Sulphuric acid, Folin -Denis reagent, sodium carbonate solution for phytochemical analysis.

2.1.3. Equipment and Apparatus

Kjeldhal Apparatus,

2.2. Methods

2.2.1. Analysis for mineral composition of *Detarium microcarpum* seeds.

Mineral composition was determined according to method of Shahidi *et al.* [27]. Air dried seeds were macerated into small component. The samples were dried in an oven at 100°C for 4 hours and then milled with an electric blender to powder form. 2 g each of samples were weighed into a cleaned porcelain crucible and subjected at 450°C in a muffle furnace to dry ash. The resultant ash was dissolved in 5ml of HNO₃/HCL/H₂O (1:2:3) and heated gently on a hot plate until brown fumes disappeared. To the remaining materials in each crucible, 5 ml of deionized water was added and heated until a colourless solution was obtained. The mineral solution in each crucible was filtered into a 100 ml volumetric and the volume made to the mark with distilled water. This solution was used for elemental analysis by atomic absorption spectrophotometer. A 10 cm long cell was used and concentration of each element in the sample was calculated on percentage of dry matter.

2.2.2. Proximate Analysis:

Air dried seeds were ground to powdered form and analyzed for their proximate compositions using the Official methods as described by AOAC, [28].

2.2.3. Determination of Phytochemical content:

Alkaloids were determined by the method described by Harbone [29]. Tannins were determined by the method of Pearson [30]. Method of Obadoni and Onyeka [31] was used to determine the saponin content while the flavonoids were determined by methods of Boham and Kocipal-Abyazam [32]. Similarly, methods of Oberlease *et al.*, [33] was adopted for Phytates composition and Oxalates was determined by the titration method.

2.2.1. Study Area

This experiment was conducted in Federal University Otuoke, Bayelsa State. The location is a secondary forest habitat which is partially or completely swampy. Otuoke is located at coordinates of 4°42'23.418"N 6°19'44.472"E.

2.2.2 Sample Collection and Identification

Fresh seeds of *Detarium microcarpum* were collected from secondary forest in Otuoke, Bayelsa State, Nigeria. The samples were taken to the Department of Biology and identified by in the Herbarium before pretreatment for Laboratory analysis.

2.2.3. Sample preparation

The collected *D. microcarpum* fruits were manually dehulled with a hammer to remove the seeds from the pericarp. The obtained seeds were air dried for three days.

2.3 Statistical analysis:

Mean and standard error were calculated for the data collected. Analysis of variance (ANOVA) was done using GENSTAT (version 12). Differences between treatment means were determined at 0.05 level of probability.

3. RESULTS

The mineral element composition of *Detarium microcarpum* seeds are presented in Table 1 showed high content of calcium, Magnesium, sodium and potassium

Akhigbe et al: Nutritional characterization of *Detarium microcarpum* Guill. & Perr., seeds used as a soup thickener in Bayelsa State.

Table 1: Mineral element composition of *Detarium microcarpum* seeds

	Mineral composition in seeds of <i>Detarium microcarpum</i>	Determined values in (mg/100 g)	Daily nutritional goals/source of goal [76,77,78]
1	Calcium	38.14 ±0.27	700-1300 (RDA) mg/day children 1000-1300(RDA) mg/day adult
2	Magnesium	57.20± 0.33	80-360(RDA) mg/day children 360-480 (RDA) mg/day adult
3	Sodium	18.21± 0.46	1200-2300 1500-2300 adult (CDRR) mg/day
4	Potassium	46.30±0.61	2000-2300 2300-3400 adult (AI) mg/day
5	Nitrogen	2.16±0.17	0.15-0.2 (RDA) g/kg/day
6	Phosphorus	4.21±0.20	460-1250 500-1250 adult (RDA) mg/day
7	Iron	0.20±0.03	7-15 children(RDA) mg/day 15-18 adult(RDA) mg/day
8	Manganese	0.16±0.05	1.8-2.3 adult (SI) mg/day
9.	Copper	0.42±0.02	1- 4 (RNI) mg/day adults
10	Zinc	0.36±0.07	3-9 (RDA) mg/day children 5-11 (RDA) mg/day adult

Values are means of triplicate analysis

Values obtain from proximate analysis of *Detarium microcarpum* seeds are presented in Table 2. The seeds have relatively low moisture content, moderate fat content and rich in proteins and carbohydrates.

Table 2: Proximate values in seeds of *Detarium microcarpum*

	Proximate parameters	Proximate values obtained from seeds(mg/100 g)	Daily nutritional goals/source of goal (78)
1	Carbohydrates	63.37± 0.32	130 g (RDA) children 130 g/d (RDA) Adults
2	Crude proteins	17.21±0.24	13-52 g/d (RDA) children 46-46 g/d (RDA) adult
3	Crude fat	5.07±0.13	25-40 g/d (AMDR) children 20-35 g/d (AMDR) adults
4	Crude ash	2.81±0.22	
5	Crude fibre	2.90±0.32	14-31 g (14g/1000k.cal) children 20-34 g (14g/1000kcal) adult
6	Moisture content	8.64±0.53	

Values are means of triplicate analysis

Results obtain from phytochemical analysis of *Detarium microcarpum* seeds are presented in Table 3. Relatively low phytochemicals were observed in this study.

Table 3. Phytochemical analysis of *Detarium microcarpum* seeds

	Phytochemicals analyzed	values obtained from (mg/100 g) of seeds
1	Alkaloids	1.50±0.36
2	Tannins	2.97±0.44
3	Flavonoids	1.22±0.67
4	Saponins	1.02±0.55
5	Oxalates	0.53±0.04
6	Phytates	0.74±0.08

Values are means of triplicate analysis



4. DISCUSSION

The mineral content of *Detarium microcarpum* seeds are shown in table 1. Potassium and sodium help to regulate body hemostasis which is essential for co-regulating ATP. Potassium is a vital cellular cation that facilitates intracellular osmotic pressure [35]. The potassium contents in seeds of *D. microcarpum* was 46.30 mg/100 g and sodium, 18.21 mg/100 g. The results are in agreement with the experiment of Ndulaka *et al.* [3] who reported potassium content of 56.82 mg/100 g in *D. microcapum* and 20.62 mg/100 g in sodium. The calcium content obtained in this study was high with a value of 38.14 mg/100 g. The findings in this study is contrary to the report of [36,37,38,39] who recorded calcium content of (0.35 g.100 g-1) in raw seeds of *D. microcapum*. Calcium is a vital mineral which forms structural component of bones and teeth. It is also facilitates muscle contraction, blood coagulation and nerve impulse transmission. Iron is also an essential constituent of cellular proteins and enzymes like hemoglobin and cytochrome P450. Iron deficiency is the major clinical cause of anaemia in females of reproductive age [40]. The iron content in seeds of *D. microcarpum* was very low 0.20 mg/100 g, this observation is in confirmation with the earlier claims of [36,37,38,39] who showed relatively lower value of 0.012 g.100 g-1. Despite the low iron content, its level in seeds of *D. microcarpum* is vital as a value of 0.031 g.100 g-1, is considered the minimum nutritional intake recommended by FAO/WHO which are 0.009 g·day-1 for males to 0.027 g·day-1 for females [46]. The magnesium contents of 57.20 mg/100 g does not correspond with the findings of Umaru *et al.* [41] who reported that pulps of the plant contain 20.5 mg/100 g magnesium. Similar report from Tchatcha [42] in Benin republic recorded lower values of (0.24 g.100 g-1). The difference in mineral composition in legumes might be due to variety or location, loamy soils retained in guinea savannah might be different from the ones found in mangrove savannah in Bayelsa State [43]. Manganese is vital for bone, nerve, brain, thyroid functioning and other cellular enzymatic reactions [44]. Manganese value recorded in this report was very low 0.16 mg/100, the experiment of Nwokocha and Nwokocha [45] who found 1.7mg/kg validates the lower levels observed in this report but contradicts the report of Balogun and Fetuga [46] who showed higher value of 195mg/kg. This result also contradicts the report of Issa *et al.* [47] whose experiment in Cotonou observed higher values of 135.54mg/kg. The variations in the contents of the different mineral elements observed in the present study, across countries could be due to the different soils compositions [47]. The low Phosphorus value of 4.21 mg/100 g obtained in this report contradicts the high value shown in the experiment of Umaru *et al.*; [41] with a value of 170 mg/100 g. The observed low Copper content of 0.42 mg/100 g validates earlier report of Oibiokpa *et al.* [48] who also observed lower values in pulp of *D. microcarpum*. Low Nitrogen content was observed in *D. microcarpum* in this research and there is a dearth of information in this regard. The proximate compositions in seeds of *D. microcarpum* revealed crude protein composition of 17.21 mg/100 g which corresponds with the report of Ndulaka *et al.* [3] who observed a protein value of 16.02% in seeds of *D. microcarpum*. The value obtained in this study was higher than that reports of Igwenyi and Akubugwo [49]. The moderate content of plant proteins in *D. microcarpum* has the potential to provide the essential amino acids required for healthy growth and repair of tissues as dietary proteins are relatively scarce [50]. Proteins are fundamental elements for metabolism of enzymes, hormones and many other molecules essential for life [51]. The crude fat content in seeds of *D. microcarpum* of 5.07 mg/100 g was close to the findings of Odenigbo and Obizoba [52]. Fats are a major source of energy and are the essential fuel for the brain and growing fetus [53]. They enhance flavour and palatability of food and make an important contribution to health. Food containing essential fatty acids that cannot be synthesized in the body are required for a range of metabolic and physiological processes to maintain the structural and functional integrity of cell membranes [54]. The percentage crude fiber in the seed 2.90 mg/100 g, contradicts the report of Akpata and Miachi [55], Barminas *et al.*, [56] who both reported higher values of crude fibre content. Fiber has many health benefits and can reduce the risk of diabetes [57] and high blood cholesterol [58]. Fiber reduces the transit time for dietary nutrients to circulate the gastrointestinal tracts, reduces low density lipoprotein and maintains gut health. Fibre rich food may modulate the digestive and absorptive process [59]. The high carbohydrate composition of the seeds 63.37 mg/100 g are comparable with the work of [60], who recorded 60.17% in *Brachystegia eurycoma* (Achi) and 51.03% in *D. microcarpum*. The result is also in line with the work of Ejiofor [61]. A high carbohydrate content is a major source of readily available energy [62, 63]. The ash content in seeds of *D. microcarpum* were also investigated and a value of 2.81 mg/100 g was recorded. The ash composition were comparable to values reported by Barminas *et al.* [56] for *Xylopiya ethiopicum* which is also used as a soup thickener. Similar reports for ash content were found, ranging between



Akhigbe et al: Nutritional characterization of *Detarium microcarpum* Guill. & Perr., seeds used as a soup thickener in Bayelsa State.

1 and 7% according to the experiment of [55, 36, 37, 64, 38]. Ash content can be used to evaluate food quality, high content of dry matter in food guarantees higher shelf-life thereby limiting microbial and pest spoilage during storage [65, 66]. The percentage moisture contents was relatively low, 8.64 mg/100 g and the value obtained in this study is in agreement with earlier work of Ndulaka *et al.* [3] who observed moisture content of 10.96%. Relatively low moisture content ranging from 2.2 to 16.7% was observed by [55, 36, 37, 64, 38]. However, the report of Uhegbu *et al.*; [36] and Amandikwa *et al.* [37] recorded higher values of (11.7-16.7%) which contradicts the finding in this study. *D. microcarpum* seeds contain some phytochemicals that can be removed or inactivated by different processing methods [35, 36, 37]. Phytate is a phytochemical that inhibits the absorption of iron, calcium, zinc, and other minerals, [67]. Although the toxicity dosage and effect of this antinutrient have not been well established or documented [68], Phytate level of 0.74 mg/100 g obtained in *D. microcarpum* seed in this study is not negligible. However, pre-treatments like soaking, fermentation and cooking before usage of the seeds can help reduce the negative effect and enhance quality of consumption [69, 70]. The Phytate (0.26%) contents reported earlier by Obun *et al.* [71] corresponds with the value obtained in the present study. However, our values are lower than 5.57% reported by Anhwange *et al.* [35]. The content of tannins in seeds of *D. microcarpum* in this study with a value of 2.97 mg/100 g contradicts the lower values obtained in the experiment of Peace and Adekunle [64], Michael *et al.* [38] who both reported a range of (0.67-0.74 mg·100 g⁻¹). This observed difference between the different results could be due to environmental conditions experienced by the seeds. Similarly, the saponin value of 1.02 mg/100 g, in this study is contrary to values (0.04-0.4 mg·100 g⁻¹) obtained in the work of Peace and Adekunle [64], Michael *et al.* [38]. Earlier findings by Tchatcha *et al.* [42] who document lower values of oxalates 0.18 mg·100 g⁻¹ is in line with the values obtained in this study with a level of 0.53 mg/100 g. Oxalates content of (0.04-0.86 mg·100 g⁻¹) according to Michael *et al.* [38] also corresponds with the findings in this report. Similarly, the flavonoids content of 1.22 mg/100 g in this report is close to the record of Peace and Adekunle [64] who recorded (0.13-1.82 mg·100 g⁻¹). Flavonoids are termed "nature's biological response modifiers" because they have been established experimentally to have the capacity to modify the body's reaction to foreign bodies. They show anti-allergic, anti-inflammatory, anti-cancerous and anti-microbial activity [72, 73]. Flavonoids are antioxidants which help to protect cells from lethal effects, free radicals and their derivatives [74]. A diet rich in phenols and flavonoids helps to boost the innate antioxidant-based defense system in the human body [74]. Alkaloids have been used as analgesics in the treatment of diseases and during surgery due to their therapeutic and pharmacological potency [75]. The alkaloids content (0.2-1.05 mg·100 g⁻¹) reported by Anhwange *et al.* [35], Uhegbu *et al.* [36], Amandikwa *et al.* [37], Peace and Adekunle [64], Michael *et al.* [44] is in agreement with the value of 1.50 mg/100 g obtained in this study. Processing methods have been known to influence the phytochemical composition of these seeds as demonstrated by Peace and Adekunle [64] and Michael *et al.* [38] on the effects of some processing methods on the phytochemical composition of *D. microcarpum* seeds, and found that roasting, soaking and boiling treatments reduced the anti-nutrients concentration in *D. microcarpum* kernels significantly.

5. CONCLUSION

The result obtained in this study reveals that *D. microcarpum* seeds have some vital nutritional components that can improve human health and thereby create a balance between economic development and poverty eradication for a healthy and sustainable ecosystem. More research is needed to bring to light the mechanism behind the physiological and biochemical effects of this legume and other crops on human health, especially in our present day where we truly need an increase in dietary supply and adequate nutritional value of food we consume.

Acknowledgements

The authors wish to thank all the staff of the Department of Plant Science and Biotechnology for their support during the period of this research.

Conflict of Interest

No conflict of interest was observed between the three Authors.

Contribution of the Authors

Mrs. Esther Omone Akhigbe and Prof. Mbosowo Etukudo were fully in charge of the sample/data analysis while Kelechi Joe-Ezeoke was in charge of sample collection and preparation. The Compilation of this paper was organized collectively by the three authors.



6. REFERENCES

- [1]. Ogwu MC, Ahana CMO and Osawaru ME. Sustainable food production in Nigeria: a case study for Bambara groundnut (*Vigna subterranea* (L.) Verdc.). *Int J Environ Sustain Dev*. 2018; 1: 68-77.
- [2]. World Health Organization (WHO). Research Guidelines for evaluating the safety and efficacy of Herbal Medicines. World health Organization regional office for the Western Pacific, Manila. 1993.
- [3]. Ndulaka JC, Ekaiko Onuh MU, Emeka F. and Okoro OA. Comparative Studies on the Nutritional and Anti – Nutritional properties of Indigenous seeds used as soup thickeners in South-East Nigeria. *Journal of Biotechnology and Biochemistry*. 2017; 3(5): 39-44.
- [4]. Gonzalez C. World poverty and food insecurity. *Penn St. J. Law And International Aff*. 3:56. 2015
- [5]. Hernandez OL, Tome AV, Gonzalez DT, Cabrera IP, Pino IY and Gort DDCG. Calculation of carbon sequestration and soil fauna associated with *Moringa oleifera* Lam. in living fences. *Centro Agricola Journal*. 2015. 42(1): 75-81.
- [6]. FAO. World Food and Agriculture – Statistical Pocketbook 2019. Rome. 2019
- [7]. Hyacinthe T, Charles P, Adama K, Diarra CS, Dicko MH, Svejgaard JJ and Diawara B. Variability of vitamins B1, B2 and minerals content in baobab (*Adansonia digitata*) leaves in East and West. *Food Sci. Nutr*. 2015; 3,17-24.
- [8]. Otori AA, Mann A. Determination of chemical composition, minerals and antinutritional factors of two wild seeds from Nupeland, North central Nigeria. *Am. J. Chem. Appl*. 2014; 1: 20-26.
- [9]. Stadlmayr B, Charrondiere UR, Eisenwagen S, Jamnadass R, Kehlenbeck K. Review: Nutrient composition of selected indigenous fruits from sub-Saharan Africa. *Journal of the science of food and Agriculture*. 2013; 93:2627-2636.
- [10]. Schreckenber K, Awono A, Degrande A, Mboosso C, Ndoye O, Tchoundjeu Z. Domesticating indigenous fruit trees as a contribution to poverty reduction. *For. trees Livelihoods*. 2006; 16:25-51.
- [11]. Ngome PIT, Shackleton C, Degrande A and Tieguhong JC. Addressing constraints in promoting wild edible plants' utilization in household nutrition: case of the Congo Basin forest area. *Agric. Food Secur*. 2017;6 (1): 20-21.
- [12]. FAO. International Scientific Symposium 'Biodiversity and Sustainable Diets', Final Document. Available on. www.fao.org/ag/humannutrition. 2013.
- [13]. Joyanes M, Lema L. Criteria for optimizing food composition tables in relation to studies of habitual food intakes. *Crit. Rev. Food Sci. Nutr*. 2006;46: 329-336.
- [14]. Ayozie CC. The effect of storage condition on the rheological properties of soup thickeners (Achi, "Ukpo", and "Ofo"). B. tech. Thesis submitted to the Dept. of Food Science and Technology, Federal University of Technology, Owerri, Nigeria. 1 – 3pp. 2010.
- [15]. Onweluzo JC, Onuoha KC, Obanu ZA. Certain functional properties of gums derived from some lesser known tropical legumes (*Azalia Africana*, *Deterium microcarpum* and *Mucuna flagellipes*) *Plant foods Hum Nutr*. 1995; 48(1):55-63
- [16]. Ezeoke C. The effect of processing on the rheological properties of some food thickeners ("Achi", "Ukpo", "Ofo"). B.tech Thesis submitted to the department of Food Science and Technology, Federal University of Technology, Owerri Nigeria. pp. 3 – 4. 2010
- [17]. Ajayi IA, Oderinde RA, Kajogbola DO, and Uponi JI. Oil content and fatty acid composition of some underutilized legumes from Nigeria. *Food chemistry*. 2006; 991 : 115 – 120.
- [18]. Adebowale BC, Lawal C. *Food Gums and Ingredients*. 2nd Edition, Academic press. 34-35pp. 1986
- [19]. Florence SP, Uroop A, Asha MR and Jyotsna Rajiv J. Sensory, Physical and Nutritional qualities of Cookies prepared from Pearl Millet (*Pennisetum typhoideum*). *Journal of food processing and technology*. 2014; 5:377
- [20]. Ruxton, C. H., Reed, S. C., Simpson, M. J. and Millington, K. J. (2004). The health benefits of omega-3 polyunsaturated fatty acids: a review of the evidence. *J Hum Nutr Diet*, 17: 449-459.

Akhigbe et al: Nutritional characterization of *Detarium microcarpum* Guill. & Perr., seeds used as a soup thickener in Bayelsa State.

- [21]. Behl T, Kumar K, Brisc C, Rus M., et al. Exploring the multifocal role of phytochemical as immunomodulators. *Biomed Pharmacotherapy*. 2021; 133: 110959.
- [22]. Tiwari S and Talreja S. Human Immune System And Importance Of Immunity Boosters On Human Body: A Review . *J. Global Trends Pharm Sci*. 2020; 11 (4): 8641 – 8649
- [23]. Khatun MAK, Matsugo S, and Konishi T. Novel Edible Mushroom BDM-X as an Immune Modulator: Possible Role in Dietary Self-Protection Against COVID-19 Pandemic. *Am J Biomed Sci & Res*. 2021; 2021 - 12(6).
- [24]. Arbonnier MA. Arbres, arbustes et lianes des zones sèches d’Afrique de l’Ouest. Montpellier, France : CIRAD-MNHN, p573. 2002
- [25]. Akah P, Nworu CS, Mbaoji FN, Nwabunike I and Onyeto CA. Genus *Detarium*: ethnomedicinal, phytochemical and pharmacological profile. *Phytopharmacology*. 2012; 2(3): 367-375
- [26]. Kouyate AM. Aspects ethnobotaniques et étude de la variabilité morphologique, biochimique et phénologique de *Detarium microcarpum* Guill. & Perr. au Mali. Thèse de doctorat : Ghent University (Belgique). 2005
- [27]. Shahidi F, Arachchi JKV and Jeon YJ. Food applications of chitin and chitosan. *Trends in food science technology*. 1999; 10; 37-51
- [28]. AOAC. Official method of Analysis. 14th Edition, Association of official analytical chemists, Washington DC.65-82pp. 1986
- [29]. Harborne JB. Text Book of phytochemical methods. A guide to modern techniques of plant analysis. 2nd edition. Chapman and Hall LTD, London.pp88-185. 1973
- [30]. Pearson D. Chemical analysis of foods. 7th edition, churchhill livingstone, London. 1976
- [31]. Obadoni BO, Onyeka PO. Phytochemical studies and comparative efficacy of the crude extracts of some haemostatic plants in Delta and Bayelsa State of Nigeria. *Global Journal of pure and applied Sciences*. 2011; 8(2):203-208
- [32]. Boham BA, Kocipal-Abyazan R. Flavonoid and condensed tannins from leaves of *Hawaiian vaccinium*, *vaticulatum* and *V. calycinlum*. *Pacific Science*. 1994; 48:458-463.
- [33]. Oberlease D, Muhere ME , O’ Dell, BL. Effects of phytic acid on zinc availability and parakeratosis in swine. *J. Anim. Sci*. 1962; 21:57-61
- [34]. Vasudevan DM, Sreekumari S. Textbook of biochemistry for medical student. Jaypee Brothers Publisher Ltd. New Delhi. pp 313-329. 2007
- [35]. Anhwange BA, Ajibola VO, Oniye SJ. Chemical studies of the seeds of *Moringa oleifera* (Lam) and *Detarium microcarpum* (Guill and Sperr). *J. Biol. Sci*. 2004; 4(6): 711-715
- [36]. Uhegbu OF, Onwuchekwa CC, Iweala EEJ and Kanu I. Effect of processing methods on nutritive and antinutritive properties of seeds of *Brachystegia eurycoma* and *Detarium microcarpum* from Nigeria. *Pak. J. Nutr*. 2009; 8(4): 316-320
- [37]. Amandikwa C, Bede EN, Eluchie CN. Effects of processing methods on proximate composition, mineral content and functional properties of "Ofor" (*Detarium microcarpum*) seed flour. *Int. J. Sci. Res*. 2017; 6(5): 966-970
- [38]. Michael K, Sogbesan OA, Onyia LU and Kefas M. Effect of processing methods on the nutritional and anti-nutritional value of *Detarium microcarpum* (Guill and Sperr) seed meals. *Int. J. Appl. Res*. 2019; 5(5): 68-72.
- [39]. Devlin TM. Textbook of Biochemistry with Clinical Correlation 6th edition. John Wiley 7 sons, Inc, Publication. pp1094-1116, 2006
- [40]. Alais C, Linden G. Food biochemistry. ellis Horwood series in food science and technology 1st ed. Springer, New York. 222p, 1991
- [41]. Umaru HA, Adamu R, Dahiru D. and Nadro MS. Levels of antinutritional factors in some wild edible fruits of Northern Nigeria. *Afr. J. Biotechnol*. 2007; 6(16):1935-1938.
- [42]. Tchatcha AD, Mureşan V, Djossou AJ, Păucean A, Tchobo FP, Mureşan A, Houndonougbo F and Soumanou MM. *Detarium microcarpum* Guill. & Perr. fruit properties, processing and food uses. A review. *Biotechnol. Agron. Soc. Environ*. 2022; 26(3):144-154.

- [43]. Abdalbasat A, Mohamed ESM, Ahmad BU. Detarium microcarpum Guill and Perr fruit proximate chemical analysis and sensory characteristics of concentrated juice and jam. *African journal of biotechnology*. 2009; 8(17):4217-4221
- [44]. Keen CL, Ensunsa JC, Watson MH, Baly DL, Donovan SM *et al.* Nutritional aspects of manganese from experimental studies, *Neurotoxicology*. 1999; 20, (2-3): 213–223.
- [45]. Nwokocha LM, Nwokocha KE. Chemical composition and rheological properties of *Detarium microcarpum* and *Irvingia gabonensis* seed flours. *Scientific African*. 2020; 10: 00529.
- [46]. Balogun M, Fetuga BL. Chemical composition of some underexploited leguminous crop seeds in Nigeria. *Journal of Agricultural and Food Chemistry*. 1986; 34(2):189– 192.
- [47]. Issa AG, Djossou AJ, Mazou M, Alitonou GA and Tchobo FP. Physicochemical Characterization of *Detarium microcarpum* Seeds from Northern Benin. *International Journal of Food Science*. 2022; 1-11
- [48]. Oibiokpa FI, Adoga GI, Saidu AN and Shittu KO. Nutritional composition of *Detarium microcarpum* fruit. *Afr. J. Food Sci*. 2014; 8(6):342-350
- [49]. Igwenyi IO, Akubugwo EI. Analysis of four seeds used as soup thickeners in the south Eastern parts of Nigeria. Conference proceeding of 2010 International Conference on Chemistry and Chemical Engineering (ICCCE, 2010), Kyoto, Japan. 426 – 430.
- [50]. Igwenyi IO. *Biochemistry: An Introductory Approach*. Willyrose and Appleseed Publishing Coy, Leach Road, Abakaliki, Ebonyi State, Nigeria 2008
- [51]. Jackson A. Protein in essentials of human nutrition, In: Mann, Jim, Stewart Truswell, A. (Eds.), second ed. Oxford University Press Inc., New York. 2002
- [52]. Odenigbo UM, Obizoba IC. Effects of food processing techniques on the nutrient and antinutrient composition of *afzelia africana* (akparata) *Journal of Biomedical Investigation*. 2004; 2(2):86-91.
- [53]. Stubbs RJ, Hopkins M, Finlayson GS, Duarte C, Gibbons C, Blundell JE. Potential effects of fat mass and fat-free mass on energy intake in different states of energy balance. *Eur. J. Clin. Nutr*. 2018; 72: 698.
- [54]. Mann J, Skeaff M. Lipids in essentials of Human Nutrition, In: Mann, Jim, Stewart Truswell, A. (Eds.), second ed. Oxford University Press Inc., New York. 2002
- [55]. Akpata MI, Miachi OE. Proximate composition and selected functional properties of *Detarium microcarpum*. *Plant Foods for Human Nutrition*. 2001; 56(4): 297– 302
- [56]. Barminas JY, James MK and Abubakar UM. Chemical composition of seeds and oil of *Xlopia aethiopica* grown in Nigeria. *Plant Foods for Human Nutrition (formerly Qualitas planetarium)*. 2004; 53(3): 193-198.
- [57]. Bazongo P, Bassole I, Nelson S, Hilou A, Dicko M, Shukla V. Characteristics, composition and oxidative stability of *Lannea microcarpa* seed and seed oil. *Molecules*. 2014; 19:2684-2693
- [58]. Liu S, Manson JE, Lee IM, Cole SR, Hennekens CR, Willett WC, Burin JE. Fruit and vegetable intake and risk of cardiovascular disease. The women's health study. *Am. J. Clin. Nutr*. 2000; 72: 922-978.
- [59]. Okaka JC, Akobundu ENT and Okaka ANC. Food and human nutrition an integrated approach. OCJ. Academic publishers, Enugu. Nigeria. pp135 – 368. 2006
- [60]. Eddy NO, Udoh CI. Proximate evaluation of the nutritional value of some soup thickeners. *Chemclass journal*. 2005; 2; 12-14
- [61]. Ejiofor MAN. Nutritional values of Ogbono (*Irvingiagabonensis var. excels*) Proc.ICRAF-IITA Conf. *Irvingiagabonensis*. Ibadan Nigeria (1994)
- [62]. Assogbadjo AE, Chadare FJ, Kakai RG, Fandohan B and Baidu-Forson JJ. Variation in biochemical composition of baobab (*Adansonia digita*) pulp, leaves and seeds in relation to soil types and tree provenances. *Agric. Ecosyst. Environ*. 2012;157: 94-99.
- [63]. Bamidele OP, Ojedokoun SO, Fasogbon BM. Physico-chemical properties of instant ogbono (*Irvingia gabonensis*) mix powder. *Food Sci. Nutr*. 2015; 3:313-318.
- [64]. Peace PD, Adekunle OA. Effects of some processing methods on antinutritional, functional and pasting characteristics of *Detarium microcarpum* seed flours. *Ann. Food Sci. Technol*. 2018; 19(1), 69-78.

Akhigbe et al: Nutritional characterization of *Detarium microcarpum* Guill. & Perr., seeds used as a soup thickener in Bayelsa State.

- [65]. Magaia T, Uamusse A, Sjöholm I, Skog K. Proximate analysis of five wild fruits of Mozambique. *Sci. World J.* 2013;1-7.
- [66]. Honfo FG, Akissoe N, Linnemann AR, Soumanou M, Van Boekel MA, Nutritional composition of shea products and chemical properties of shea butter: a review. *Crit. Rev. Food Sci. Nutr.* 2014; 54: 673-686.
- [67]. Fergusin EL, Gibson RA, Opara-obisaw O, Stephen AM and Thomson LU. The zinc, calcium, copper, manganese, nonstarch polysaccharide and phytate content of seventy eight locally grown and prepared African foods,” *Journal of Food Analysis.* 1993; 6(1): 87-99.
- [68]. Ekpa E, Sani D. Phytochemicals and anti-nutritional studies on some commonly consumed fruits in Ilokoja, Kogi State of Nigeria. Open access text. 2018;
- [69]. Lestienne L. Contribution à l'étude de la biodisponibilité du fer et du zinc dans le grain de mil et conditions d'amélioration dans les aliments de complément,” in Thèse, p. 303, Université de Montpellier II, Montpellier. 2004
- [70]. Esenwah CN, Ikenebomeh MJ. Processing effects on the nutritional and anti-nutritional contents of African locust bean (*Parkia biglobosa* Benth.) seed,” *Pakistan Journal of Nutrition.* 2008; 7(2): 214-217.
- [71]. Obun CO, Yahaya MS, Kibon A and Ukim C. Effect of dietary inclusion of raw *Detarium microcarpum* seed meal on the performance and carcass and organ weights of broiler chicks. *American Journal of Food and Nutrition.* 2011;1(3):128-135.
- [72]. Yamamoto Y, Gaynor, R. Therapeutic potential of inhibition of the NF-KappaB pathway in the treatment of inflammation and cancer. *Journal of clinical investigation.* 2006
- [73]. Cushine TP, Lamb AJ. Antimicrobial activity of flavonoids. *International Journal of antimicrobial agents.* 2005; 26:343-356
- [74]. Tonukari N, Tonukari N, Ezedom T, Enuma C, Sakpa S, Avwioroko O, Eraga L and Odiyoma E. White Gold: cassava as an industry base. *American Journal of plant Sciences.* 2015; 6:972-979
- [75]. Mothes W, Heinrich U, Graf R, Nilsson I, Heijne V, Brunner J, Rapoport A. Molecular mechanism of membrane protein integration into the endoplasmic reticulum. *Cell.* 1997; 4(89): 523-533
- [76]. FAO. Human vitamins and mineral requirements. Rome, pp299-300. 2001
- [77]. Sousa C, Moutinho C, Vinha AF, Matos C. Trace minerals in human health: iron, zinc, copper, manganese and fluorine. *International journal of science and research methodology.* 2019;13(3);2024.
- [78]. USDA. Dietary guidelines for Americans 2020-2025.pp133